Vol 25, No. 1 (2024) http://www.veterinaria.org

Article Received: Revised: Accepted:



Syringaldehyde Attenuates Chronic Constriction Injury-Induced Neuropathic Pain In Sprague-Dawley Rats Via PGE2-iNOS Inhibition

Annasaheb S. Kalange¹, Murugan Vedigounder², Geetha K Mukundan³*

^{1,2,3*}Department Of Pharmacology, College of Pharmaceutical Sciences, Dayananda Sagar University, Bangalore 560078, Karnataka, India.

*Corresponding author: Geetha K Mukundan

*Department Of Pharmacology, College of Pharmaceutical Sciences, Dayananda Sagar University, Bangalore 560078, Karnataka, India.

Abstract

Neuropathic pain is a one of the most unbearable pain condition severely affecting day-to-day life in affected patients. Syringaldehyde is a compound isolated from various plant species. Syringaldehyde has shown several pharmacological properties including anti-inflammatory, anti-oxidant activities. In this work, to assess in-vivo pharmacological effect of Syringaldehyde in development and maintainace of neuropathic pain, male SD rats were subjected to CCI ligation surgery of sciatic nerve with assessment of mechanical allodynia using von Frey filaments and cold allodynia using acetone test from post-surgery day 3 onwards. CCI rats were given Syringaldehyde orally from day 7 to 14 and pain responses were recorded on day 7, 14. At the end of study, expression levels of PGE2 and iNOS mRNA was assessed in sciatic nerve of CCI rats. CCI surgery in rats developed mechanical and cold allodynia from post-surgery day 3 onwards with peak allodynia observed at day 7, and allodynia was maintained till day 14. Treatment with Syringaldehyde for from day 7 to day 14 showed reversal of mechanical and cold allodynia inhibition of CCI rats when allodynia was assessed on day 7 and day 14. Moreover, treatment with Syringaldehyde decreased mRNA expression levels of PGE2 and iNOS in sciatic nerve. The results from this study demonstrated the analgesic potential of Syringaldehyde in neuropathic pain induced by CCI surgery which may be mediated via inhibition of PGE2 and iNOS.

Keywords: CCI, Syringaldehyde, PGE2, iNOS, Allodynia, Neuropathic pain

Introduction

Neuropathic pain affects peripheral or central nervous systems and is a chronic, debilitating pain condition (Singh et al., 2017, Bernetti et al., 2021, Kaur et al., 2016). Nerve injury may occur as a result of various conditions like trauma, infection, inflammation, tumors, metabolic or endocrine diseases (Baron, 2006). Nerve injury causes inflammation at site of injury, activates nociceptors (De Jongh et al., 2003) which develops hyperalgesia (increased pain sensitivity to noxious stimuli) or allodynia (pain sensitivity to non-noxious stimuli) (Zanjani et al., 2006; Kaur et al., 2016). The available treatments for chronic neuropathic pain have limited efficacy in most patients (Finnerup et al., 2010). Thus, there is an unmet need to discover better therapies for management of neuropathic pain. Presently, treatment of neuropathic pain in patients include use of antiepilpetics, SSRI inhibitors, sodium and calcium channel blockers, NMDA antagonists; however utility of these treatment options is limited due to CNS adverse effects (Xu et al., 2012). Thus, management of neuropathic pain therapy remains an unmet medical need and it is necessary to focus on discovery of new therapeutic options for treatment of pain.

Plant derived phytochemical compounds are potential candidates for discovery of novel compounds for treatment of pain (Kumar et al., 2015). Syringaldehyde is a natural flavonoid isolated from various plants like Magnolia officinalis (Shen et al., 2009). There are different pharmacological properties exhibited by Syringaldehyde including anti-inflammatory, anti-oxidant, cardioprotective, neuroprotective and anti-diabetic activity (Stanikunaite et al., 2009; Bozkurt et al., 2014; Wu et al., 2022). Thus, Syringaldehyde can be potential therapeutic option in the treatment of neuropathic pain.

The present study aimed to ascertain pharmacological activity of Syringaldehyde in rat model of chronic constriction injury (CCI) induced neuropathic pain. To assess influence of biomarker in neuropathic pain, expression levels of pain biomarker was measured in sciatic nerve of rats with CCI injury.

Material and methods

Animals

Animal house of Dayananda Sagar University (Bengaluru, India) supplied adult male Sprague-Dawley rats (180- 200 g, 7-8 weeks old). All animals were maintained under controlled conditions like 12-h light-dark cycle and free access to food, water. Before experimental activity, animals were acclimatized to laboratory conditions for one week. Dayananda Sagar University's Institutional Animal Ethics Committee (IAEC) approved the experimental methods and procedures in animals.

REDVET - Revista electrónica de Veterinaria - ISSN 1695-7504

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Compounds and treatment

Syringaldehyde and Gabapentin were procured from Sigma (Sigma, India). Syringaldehyde (5, 15, 50 mg/kg, po) and Gabapentin were given orally to rats in neuropathic pain model of CCI.

CCI surgery for induction of neuropathic pain in rats

Neuropathic pain was induced by CCI surgery of sciatic nerve as per procedure described earlier (Bennett and Xie, 1988). Rats were anesthetized with ketamine:xylazine mixture (80:10 mg/kg, i.p.), an incicsion was given near left thigh region, bluntly dissected biceps femoris muscle to expose left sciatic nerve. Sciatic nerve was separated from connective tissue and ligated with 4 loose knots using 4-0 chromic gut sutures. The overlying muscle and skin wound was closed using silk suture 3-0.

Recording Mechanical allodynia

A set of 8 von Frey filaments (Bioseb) ranging from 0.4 g to 15 g was used to measure mechanical paw withdrawal threshold (PWT) in rats by up-down technique (Chaplan et al., 1994). Baseline PWT was measured before CCI surgery and development of allodynia was assessed by measuring PWT on day 3, 6 post surgery. On day 7, rats with mechanical allodynia were divided into 5 groups; vehicle control, Syringaldehyde (5, 15, 50 mg/kg) and Gabapentin (100 mg/kg). Treatment was given for 8 days from day 7 to day 14 surgery and effect of treatment was assessed by recording PWT on day 7 and day 14 at 1 h post treatment. Effect of compounds was ascertained by calculating percentage maximum possible effect (%MPE) using formula:

%MPE= (post-treatment PWT – vehicle PWT)/(15 - vehicle PWT) x 100

Recording Cold allodynia (Acetone test)

The acetone test for recording cold allodynia was done following procedure described earlier (Choi et al., 1994). Briefly, rats were acclimated to test environment on wire mesh floor and acetone bubble was applied to the plantar surface of the hind paw. Paw withdrawal latency (PWL) for response (flinching of hindpaw) was recorded till 30 seconds following acetone application. Baseline PWL was measured before CCI surgery and development of allodynia was assessed by measuring PWL on day 3, 6 post surgery. On day 7, rats were divided into 5 groups; vehicle control, Syringaldehyde (5, 15, 50 mg/kg) and Gabapentin (100 mg/kg). One-hour after oral administration, effect of treatment was assessed by recording PWL. Treatment was continued for 8 days till day 14 surgery and effect of treatment was assessed by recording PWL on day 10 and day 14. Effect of compounds was ascertained by calculating percentage maximum possible effect (%MPE) using formula:

%MPE = (post-treatment PWL - vehicle PWL)/(30 - vehicle PWL) x 100

mRNA analysis

PGE2 and iNOS mRNA expression levels were measured in CCI operated sciatic nerve samples using reverse transcription quantitative polymerase chain reaction (RT-PCR) assay. After recording of PWL/ PWT, animals were sacrificed and ipsilateral (left CCI operated) sciatic nerve samples were collected to assess mRNA expression. Contralateral (right CCI non-operated) sciatic nerve samples from vehicle group were collected and used as normal control sciatic nerve. Sciatic nerve samples were homegenised, total RNA was extracted using RNA extraction kit (Krishgen Biosystems). After measuring RNA concentration and purity, isolated RNA was reverse transcribed into single stranded cDNA using iScript cDNA synthesis kit (Bio-Rad). RT-PCR was performed with SYBR green (Bio-Rad) using RT-PCR detection system (Applied Biosystems). Primers used for gene expression are mentioned in Table 1. Gene expression levels of pain mediators in test samples was normalized with GAPDH (housekeeping gene) and relative fold over gene expression was calculated as -

Relative expression= 2^-(Ct GAPDH – Ct target gene)

Table 1. List of primers

Gene	Primer sequence
GAPDH	F-5'- ATGCTGGTGCTGAGTATGTC -3' & R-5'- AGTTGTCATATTTCTCGTGGGTT -3'
PGE2	F-5'-CCCACTCACCTGCTGCTACTC-3' & R-5'-AGAAGTGCTTGAGGTGGTTGTG-3'
iNOS	F-5'-CGAAACGCTTCACTTCCAA-3' &R-5'-TGAGCCTATATTGCTGTGGCT-3'

Statistical analysis

To analyse statistical outcomes of study, 2-way analysis of variance (ANOVA) was performed with Bonferroni post-test and 1-way ANOVA was performed with Dunnett's post-test as specified in data.

Results

Induction of allodynia post surgery

Prior to CCI surgery, baseline PWT and PWL were recorded in rats and rats were subjected to CCI surgery. Development of mechanical allodynia was seen from day 3 onwards, as PWT of rats decreased from 11.4 g to 1.9 g on day 3. Mechanical

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allodynia was maintained till day 14. Similarly, in response to CCI surgery, cold allodynia was seen as PWL of rats decreased from 29.9 seconds to 7.8 seconds on day 3. As seen in mechanical allodynia, cold allodynia was also maintained till day 14 post surgery (table 2).

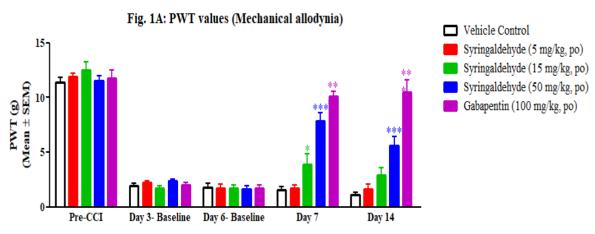
Table 2. Induction of mechanical and cold allodynia

Allodynia	Surgery day				
Allodynia	Pre-CCI	Day 3	Day 6	Day 7	Day 14
Mechanical allodynia (PWT in grams)	11.4	1.9	1.8	1.6	1.1
Cold allodynia (PWL in seconds)	29.9	7.8	7.1	7.4	6.2

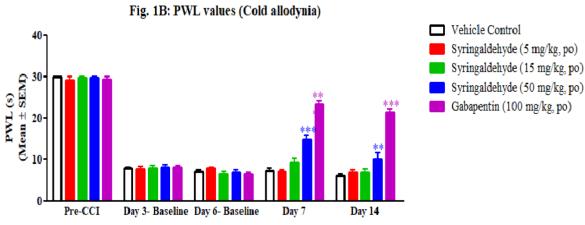
Effect of Syringaldehyde treatment on CCI induced mechanical and cold allodynia

Post recording of baseline PWT and PWL on day 6, rats were randomized into 5 groups as per day 6 baseline mechanical allodynia. On day 7, rats were treated with vehicle/ Syringaldehyde/ Gabapentin and response was recorded at 1 h post treatment. Syringaldehyde treatment increased PWT (figure 1A) and PWL (figure 1B). Significant analgesic effect in terms of %MPE was seen with Syringaldehyde at dose of 15 mg/kg (P<0.05) and 50 mg/kg (P<0.001) for mechanical allodynia (figure 2A), while for cold allodynia 50 mg/kg dose showed significant analgesic effect (P<0.001, figure 2B). As expected, positive control Gabapentin showed significant analgesic effect for mechanical and cold allodynia (P<0.001). Treatment with Syringaldehyde was continued till day 14, and effect of treatment was assessed on day 14 at 1 h post treatment. Significant analgesic effect in terms of %MPE was seen with Syringaldehyde at dose 50 mg/kg for mechanical (P<0.001, figure 2A) as well as cold allodynia (P<0.01, figure 2B). As expected, positive control Gabapentin showed significant analgesic effect for mechanical and cold allodynia (P<0.001).

Figure 1. Effect of Syringaldehyde on CCI induced neuropathic pain. Paw withdrawal threshold (PWT) and paw withdrawal latency (PWL) values are expressed in as Mean ± SEM



^{***}P<0.001, *P<0.05 vs Vehicle Control, Two-way ANOVA followed by Bonferroni post test



***P<0.001, **P<0.01 vs Vehicle Control, Two-way ANOVA followed by Bonferroni post test

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Figure 2. Effect of Syringaldehyde on CCI induced neuropathic pain. % MPE values are expressed in as Mean ± SEM

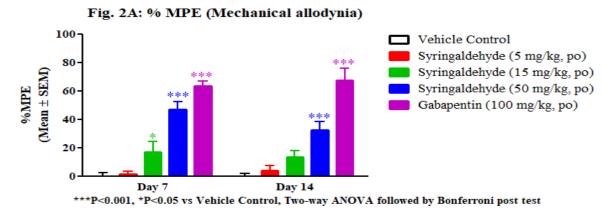
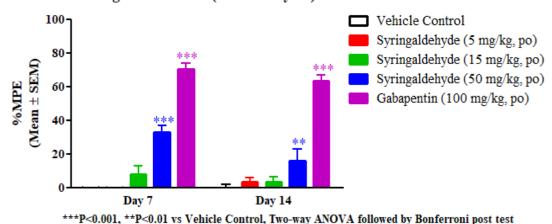


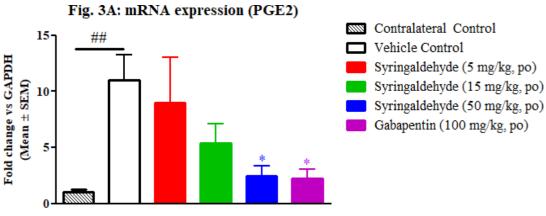
Fig. 2B: % MPE (Cold allodynia)



Effect of Syringaldehyde treatment on CCI induced biomarker expression levels

CCI surgery increased mRNA expression of PGE2 (11.03 fold, P<0.01, fig. 3A) and iNOS (12.97 fold, P<0.05, fig. 3B) in ipsilateral hind paw as compared to contralateral hind paw. Treatment with Syringaldehyde (5, 15, 50 mg/kg) decreased PGE2 mRNA levels (9.01, 5.40, 2.43 fold respectively, fig. 3A) and iNOS mRNA levels (10.17, 6.96 and 2.92 fold respectively, fig. 3B) with 50 mg/kg dose showing statistically significant reversal of PGE2 mRNA (P<0.01) and iNOS mRNA (P<0.05).

Figure 3. Effect of Syringaldehyde on CCI induced neuropathic pain. Gene expression fold over vs GAPDH values are expressed in as Mean \pm SEM



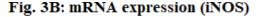
*P<0.05 vs Vehicle Control, ##P<0.01, One-way ANOVA followed by Dunnett's post test

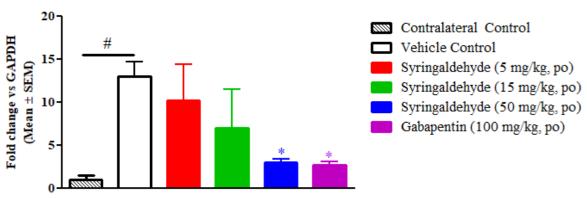
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*P<0.05 vs Vehicle Control, #P<0.05, One-way ANOVA followed by Dunnett's post test

Discussion

CCI induced neuropathic pain is the most a widely used animal model to study mechanisms of neuropathic pain and CCI surgery model mimic pathophysiology of complex regional pain syndrome in humans (Bennet and Xie 1988, Jin et al., 2008). Sciatic nerve injury leads to local nerve inflammation, alters nerve structure and function which causes increase in pain sensitivity (Jaggi and Singh, 2010). CCI surgery causes an injury to the sensory afferents nerve fibers and develops long-term allodynia post-surgery (Bennet and Xie 1988, Austin et al., 2012). Nociceptors innervated with C fibres and thinly myelinated Ad fibres are physiologically involved in pain signalling. However, nerve injury results in phenotypic switch where involvement of Aß fibers in pain signalling leads to increased pain sensitivity and allodynia (Basbaum and Fields, 1984; Mahmoud et al., 2021). Therefore, synaptic transmission in the spinal cord is enhanced, and the response to innocuous stimuli is exaggerated (Neumann et al., 1996). In conformance with literature reports (Bennet and Xie, 1988; Pradhan et al, 2010), data from this study demonstrated that CCI surgery resulted in development of mechanical and cold allodynia in rats from day 3 post CCI surgery. Acute oral treatment of Syringaldehyde at dose of 5, 15, 50 mg/kg on day 7 post surgery has exhibited significant efficacy at 1 h post-treatment. Additionally, 8 day chronic dosing of Syringaldehyde from day 7 to day 14 significantly alleviated allodynia which was assessed on day 10 and day 14. As noted in literature reports, treatment with Gabapentin reversed allodynia in CCI model of neuropathic pain (Rutten et al., 2018).

Biomarker analysis of sciatic nerve showed that increased expression of PGE2, iNOS mRNA levels was seen in ipsilateral sciatic nerve with CCI surgery. Literature reports indicate higher levels of pro-inflammatory pain mediators like COX-2 and PGE2 due to CCI surgery of sciatic nerve in rats (Rezq et al., 2020). Higher expression of COX-2 leads to increased release of PGE2 and higher PGE2 levels increases pain sensitivity and develops neuropathic pain (Ma and Quirion 2005; Mahmoud et al., 2021). Similarly, in this study, CCI surgery led to increased mRNA levels of PGE2 in sciatic nerve at 14 days post-surgery treatment and treatment with Syringaldehyde dose dependently reversed increased levels of these pro-inflammatory pain mediators. Furthermore, CCI induced neuropathic pain is accompanied with increased gene expression levels of iNOS in sciatic nerve (Sobeh et al., 2020). Higher expression of iNOS is known to drive development of neuropathic pain following peripheral nerve injury (Sobeh et al., 2019). At the site of nerve injury, iNOS activation triggers Nitric oxide (NO) release that modulates inflammatory processes and mediates central mechanisms in development of allodynia (Meller et al., 1992). Limiting NO release by inhibition of iNOS is a viable strategy in management of pain (Korhonen et al. 2005.; Gao et al., 2012). Similarly, in this study, chronic treatment with Syringaldehyde reduced elevalted iNOS mRNA levels. Attenuation of neuropathic pain by Syringaldehyde is probably attributed to inhibition of PGE2, iNOS in sciatic nerve.

Summarily, this study demonstrates antiallodynic effects of Syringaldehyde in CCI induced neuropathic pain. Chronic treatment of Syringaldehyde was effective in reversing CCI induced neuropathic pain in rats and inhibited pain biomarkers increased in response to CCI in rats. Thus, going further, Syringaldehyde can be a potential novel therapy for management of neuropathic pain conditions.

Conclusion

This study has demonstrated pharmacological effect of Syringaldehyde in CCI induced neuropathic pain model. Treatment with Syringaldehyde modulates expression of pain mediators to exert its pharmacological effect in CCI induced neuropathic pain model. Data from this study, demonstrates potential of Syringaldehyde as a novel therapeutic option with pharmacological efficacy for treatment of pain.

REDVET - Revista electrónica de Veterinaria - ISSN 1695-7504

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http://www.veterinaria.org

Article Received: Revised: Accepted:



Conflict of interest

The authors have no conflicts of interest regarding this investigation.

References

- 1. Austin PJ, Wu A, Moalem-Taylor G. Chronic constriction of the sciatic nerve and pain hypersensitivity testing in rats. J Vis Exp. 2012 Mar 13;(61):3393. doi: 10.3791/3393.
- 2. Baron R. Mechanisms of disease: neuropathic pain--a clinical perspective. Nat Clin Pract Neurol. 2006 Feb;2(2):95-106. doi: 10.1038/ncpneuro0113.
- 3. Basbaum AI, Fields HL. Endogenous pain control systems: brainstem spinal pathways and endorphin circuitry. Annu Rev Neurosci. 1984;7:309-38. doi: 10.1146/annurev.ne.07.0301 b84.001521.
- 4. Bennett GJ, Xie YK. A peripheral mononeuropathy in rat that produces disorders of pain sensation like those seen in man. Pain. 1988 Apr;33(1):87-107. doi: 10.1016/0304-3959(88)90209-6.
- 5. Bennett, G. J., and Xie, Y.-K. (1988). A peripheral mononeuropathy in rat that produces disorders of pain sensation like those seen in man. Pain 33, 87–107. doi: 10.1016/0304-3959(88)90209-6
- 6. Bernetti A., Agostini F., de Sire A., Mangone M., Tognolo L., Di Cesare A., et al. (2021). Neuropathic Pain and Rehabilitation: A Systematic Review of International Guidelines. Diagnostics 11 (1), 74. Doi-10.3390/diagnostics11010074
- 7. Bozkurt AA, Mustafa G, Tarık A, Adile O, Murat SH, Mesut K, Yıldıray K, Coskun S, Murat C. Syringaldehyde exerts neuroprotective effect on cerebral ischemia injury in rats through anti-oxidative and anti-apoptotic properties. Neural Regen Res. 2014 Nov 1;9(21):1884-90. doi: 10.4103/1673-5374.145353.
- 8. Chaplan S. R., Bach F. W., Pogrel J. W., Chung J. M., Yaksh T. L. (1994). Quantitative assessment of tactile allodynia in the rat paw. J. Neurosci. Methods 53, 55–63. 10.1016/0165-0270(94)90144-9
- 9. Choi Y, Yoon YW, Na HS, Kim SH, Chung JM (1994). Behavioral signs of ongoing pain and cold allodynia in a rat model of neuropathic pain. Pain, 59 (3): 369-376. DOI: 10.1016/0304-3959(94)90023-X
- 10. De Jongh RF, Vissers KC, Meert TF, Booij LHDJ, De Deyne CS, Heylen RJ. The role of interleukin-6 in nociception and pain. Anesth Analg. 2003 Apr;96(4):1096-1103. doi: 10.1213/01.
- 11. Finnerup NB, Sindrup SH, Jensen TS. The evidence for pharmacological treatment of neuropathic pain. Pain. 2010 Sep;150(3):573-581. doi: 10.1016/j.pain.2010.06.019.
- 12. Gao Y, Jiang W, Dong C, Li C, Fu X, Min L, Tian J, Jin H, Shen J. Anti-inflammatory effects of sophocarpine in LPS-induced RAW 264.7 cells via NF-kB and MAPKs signaling pathways. Toxicol In Vitro. 2012 Feb;26(1):1-6. doi: 10.1016/j.tiv.2011.09.019.
- 13. Jaggi AS, Singh N. Differential effect of spironolactone in chronic constriction injury and vincristine-induced neuropathic pain in rats. Eur J Pharmacol. 2010 Dec 1;648(1-3):102-9. doi: 10.1016/j.ejphar.2010.08.050.
- 14. Jin Y, Sato J, Yamazaki M, Omura S, Funakubo M, Senoo S, Aoyama M, Mizumura K. Changes in cardiovascular parameters and plasma norepinephrine level in rats after chronic constriction injury on the sciatic nerve. Pain. 2008 Apr;135(3):221-231. doi: 10.1016/j.pain.2007.05.020.
- 15. Kaur, G., Bedi, O., Sharma, N., Singh, S., Deshmukh, R. and Kumar, P. (2016) Anti-hyperalgesic and anti-nociceptive potentials of standardized grape seed proanthocyanidin extract against CCI-induced neuropathic pain in rats. Journal of Basic and Clinical Physiology and Pharmacology, Vol. 27 (Issue 1), pp. 9-17. https://doi.org/10.1515/jbcpp-2015-0026
- 16. Korhonen R, Lahti A, Kankaanranta H, Moilanen E. Nitric oxide production and signaling in inflammation. Curr Drug Targets Inflamm Allergy. 2005 Aug;4(4):471-9. doi: 10.2174/1568010054526359.
- 17. Kumar A, Agarwal K, Maurya AK, Shanker K, Bushra U, Tandon S, Bawankule DU. Pharmacological and phytochemical evaluation of Ocimum sanctum root extracts for its antiinflammatory, analgesic and antipyretic activities. Pharmacogn Mag. 2015 May;11(Suppl 1):S217-24. doi: 10.4103/0973-1296.157743.
- 18. Ma, W. and Quirion, R. (2005), Up-regulation of interleukin-6 induced by prostaglandin E2 from invading macrophages following nerve injury: an in vivo and in vitro study. Journal of Neurochemistry, 93: 664-673. https://doi.org/10.1111/j.1471-4159.2005.03050.x
- 19. Mahmoud MF, Rezq S, Alsemeh AE, Abdelfattah MAO, El-Shazly AM, Daoud R, El Raey MA and Sobeh M (2021) Potamogeton perfoliatus L. Extract Attenuates Neuroinflammation and Neuropathic Pain in Sciatic Nerve Chronic Constriction Injury-Induced Peripheral Neuropathy in Rats. Front. Pharmacol. 12:799444. doi: 10.3389/fphar.2021.799444
- 20. Mahmoud MF, Rezq S, Alsemeh AE, Abdelfattah MAO, El-Shazly AM, Daoud R, El Raey MA and Sobeh M (2021) Potamogeton perfoliatus L. Extract Attenuates Neuroinflammation and Neuropathic Pain in Sciatic Nerve Chronic Constriction Injury-Induced Peripheral Neuropathy in Rats. Front. Pharmacol. 12:799444. doi: 10.3389/fphar.2021.799444
- 21. Meller ST, Pechman PS, Gebhart GF, Maves TJ. Nitric oxide mediates the thermal hyperalgesia produced in a model of neuropathic pain in the rat. Neuroscience. 1992 Sep;50(1):7-10. doi: 10.1016/0306-4522(92)90377-e.
- 22. Pradhan AA, Yu XH, Laird JM. Modality of hyperalgesia tested, not type of nerve damage, predicts pharmacological sensitivity in rat models of neuropathic pain. Eur J Pain. 2010 May;14(5):503-9. doi: 10.1016/j.ejpain.2009.08.010.

REDVET - Revista electrónica de Veterinaria - ISSN 1695-7504

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http://www.veterinaria.org

Article Received: Revised: Accepted:



- 23. Rezq, S., Alsemeh, A.E., D'Elia, L. et al. Thymus algeriensis and Thymus fontanesii exert neuroprotective effect against chronic constriction injury-induced neuropathic pain in rats. Sci Rep 10, 20559 (2020). https://doi.org/10.1038/s41598-020-77424-0
- 24. Rutten K, Gould SA, Bryden L, Doods H, Christoph T, Pekcec A. Standard analgesics reverse burrowing deficits in a rat CCI model of neuropathic pain, but not in models of type 1 and type 2 diabetes-induced neuropathic pain. Behav Brain Res. 2018 Sep 17;350:129-138. doi: 10.1016/j.bbr.2018.04.049.
- 25. Saliba S. W., Jauch H., Gargouri B., Keil A., Hurrle T., Volz N., Mohr F., Stelt M., Bräse S., Fiebich B. L. (2018): Anti-neuroinflammatory effects of GPR55 antagonists in LPS-activated primary microglial cells. J. Neuroinflammation. 15, 322.
- 26. Singh H., Bhushan S., Arora R., Singh Buttar H., Arora S., Singh B. (2017). Alternative Treatment Strategies for Neuropathic Pain: Role of Indian Medicinal Plants and Compounds of Plant Origin-A Review. Biomed. Pharmacother. 92, 634–650. Doi-10.1016/j.biopha.2017.05.079
- 27. Sobeh M, Mahmoud MF, Rezq S, Abdelfattah MAO, Mostafa I, Alsemeh AE, El-Shazly AM, Yasri A, Wink M. Haematoxylon campechianum Extract Ameliorates Neuropathic Pain via Inhibition of NF- KB/TNF-a/NOX/INOS Signalling Pathway in a Rat Model of Chronic Constriction Injury. Biomolecules. 2020; 10(3):386. https://doi.org/10.3390/biom10030386
- 28. Sobeh M, Mahmoud MF, Rezq S, Alsemeh AE, Sabry OM, Mostafa I, Abdelfattah MAO, Ait El-Allem K, El-Shazly AM, Yasri A, et al. Salix tetrasperma Roxb. Extract Alleviates Neuropathic Pain in Rats via Modulation of the NF-kB/TNF-a/NOX/iNOS Pathway. Antioxidants. 2019; 8(10):482. https://doi.org/10.3390/antiox8100482
- 29. Stanikunaite R, Khan SI, Trappe JM, Ross SA. Cyclooxygenase-2 inhibitory and antioxidant compounds from the truffle Elaphomyces granulatus. Phytother Res. 2009 Apr;23(4):575-8. doi: 10.1002/ptr.2698.
- 30. Wu J, Fu YS, Lin K, Huang X, Chen YJ, Lai D, Kang N, Huang L, Weng CF. A narrative review: The pharmaceutical evolution of phenolic syringaldehyde. Biomed Pharmacother. 2022 Sep;153:113339. doi: 10.1016/j.biopha.2022.113339.
- 31. Xu B, Descalzi G, Ye H-R, Zhuo M, Wang Y-W. Translational Investigation and Treatment of Neuropathic Pain. Molecular Pain. 2012;8. doi:10.1186/1744-8069-8-15
- 32. Zanjani TM, Sabetkasaei M, Mosaffa N, Manaheji H, Labibi F, Farokhi B. Suppression of interleukin-6 by minocycline in a rat model of neuropathic pain. Eur J Pharmacol. 2006 May 24;538(1-3):66-72. doi: 10.1016/j.ejphar.2006.03.063.