http://www.veterinaria.org

Article Received: Revised: Accepted:



In-Silico Studies On Astilbin As A Potential Antioxidant Agent: A Multi-Faceted Computational Approach

Pratikkumar Gavval^{1*}, Shrinivas Mohite², Sandeep Kane³, Sonali Tambave⁴, Pallavi Sutar⁵, Vidya Dange⁶, Avani Shewale⁷, Shailaja Kamble⁸

1*,2,3,4,5 Department of Pharmaceutical Chemistry, Rajarambapu College of Pharmacy Kasegaon.
 6Department of Pharmaceutics, Rajarambapu College of Pharmacy Kasegaon.
 7.8 Department of Pharmacology, Rajarambapu College of Pharmacy Kasegaon.

*Corresponding Author: Pratikkumar Gavva

*Department of Pharmaceutical Chemistry, Rajarambapu College of Pharmacy Kasegaon. Tal-Walwa Dist-Sangli, Maharashtra, India.

Abstract

Astilbin, a prominent flavonoid derived from the rhizomes of Smilax glabra, exhibits significant antioxidant properties. This study employs a comprehensive in-silico approach, encompassing Swiss Similarity studies, ADME and toxicity predictions, Cytoscape network analysis, and molecular docking studies, to elucidate the potential of astilbin as an effective antioxidant agent. Swiss Similarity analysis identified structurally similar compounds, facilitating the selection of promising candidates. ADME and toxicity predictions confirmed favourable pharmacokinetic properties and low toxicity. Cytoscape network analysis highlighted key protein interactions, and molecular docking studies demonstrated strong binding affinities of astilbin to antioxidant-related enzymes, with comparative analysis against the standard antioxidant ascorbic acid. These findings underscore the potential of astilbin in antioxidant therapy, warranting further experimental validation.

Keywords: Astilbin, Molecular Docking, Swiss Similarity, ADME, Toxicity Predictions.

Introduction

Natural compounds have been extensively studied for their potential therapeutic properties, particularly as antioxidants. Antioxidants are crucial in mitigating oxidative stress, which is implicated in various chronic diseases, including cancer, cardiovascular diseases, and neurodegenerative disorders. Among these natural compounds, astilbin, a flavonoid extracted from the rhizomes of Smilax glabra, has shown promising antioxidant activity.

Astilbin has been traditionally used in Chinese medicine for its anti-inflammatory, hepatoprotective, and immunomodulatory effects. Recent studies have suggested that astilbin's antioxidant properties could be attributed to its ability to scavenge free radicals and upregulate endogenous antioxidant defenses. This study aims to explore the antioxidant potential of astilbin using in-silico methods, providing a detailed analysis of its pharmacokinetic properties, toxicity, protein interactions, and binding affinities.

Materials and Methods

Swiss Similarity Studies

The Swiss Similarity platform (http://www.swisssimilarity.ch) was utilized to identify compounds structurally similar to astilbin. This tool integrates data from major chemical databases such as PubChem, ChemSpider, and ZINC, allowing for comprehensive molecular similarity searches and virtual screening. The identification of similar compounds aids in predicting potential bioactivity and discovering new therapeutic candidates.

ADME and Toxicity Predictions

ADME (Absorption, Distribution, Metabolism, and Excretion) and toxicity properties of astilbin were predicted using ADMET LAB 3.0 (http://admetlab.org). This tool evaluates critical pharmacokinetic parameters, including bioavailability, half-life, clearance rates, and tissue distribution, as well as toxicity profiles to ensure the compound's safety and efficacy.

Cytoscape Network Analysis

Cytoscape (http://www.cytoscape.org) was employed for visualizing protein-protein interactions and pathway analysis. Data from STRING (http://string-db.org) and KEGG (http://www.genome.jp/kegg) databases were integrated to create detailed interaction maps, highlighting pathways involved in oxidative stress and antioxidant activity. This analysis identified potential drug targets and elucidated the molecular mechanisms underlying astilbin's antioxidant effects.

Molecular Docking Studies

Molecular docking studies were conducted using Molsoft's ICM PRO

http://www.veterinaria.org

Article Received: Revised: Accepted:



(https://www.molsoft.com/icm_pro.html). Proteins selected for docking studies were identified through Cytoscape network analysis, focusing on key antioxidant enzymes such as Human Erythrocyte Catalase (PDB ID: 1DGF) and Human Peroxiredoxin 3 (PDB ID: 5UCX). The binding affinities and interaction mechanisms of astilbin were compared against the standard antioxidant ascorbic acid to evaluate its relative efficacy.

Results

Swiss Similarity Studies

The Swiss Similarity analysis identified several compounds with significant structural similarity to astilbin. These compounds were further analyzed to assess their potential antioxidant activity. The top-ranked compounds showed high similarity scores, indicating a strong likelihood of shared biological activity.

Table 1: Top Structurally Similar Compounds Identified in Swiss Similarity Analysis

SR.NO	MOLECULAR FORMULA	NAME	SMILES
1.	C ₁₅ H ₁₂ O ₇	2-(3,4-Dihydroxyphenyl)-3,5,7- trihydroxy-2,3-dihydro-4H-chromen-4- one	c1cc(c(cc1C2C(C(=O)c3c(cc(cc3O2)O)O)O)O)O
2.	C ₁₅ H ₁₂ O ₇	(2R,3R)-2-(3,4-Dihydroxyphenyl)-3,5,7-trihydroxy-2,3-dihydro-4H-chromen-4-one	c1cc(c(cc1[C@@H]2[C@H](C(=O)c3c(cc(cc3O 2)O)O)O)O)O
3.	C16H14O7	(2S,3S)-2-(3,4-Dihydroxyphenyl)-3,5,7-trihydroxy-6-methyl-2,3-dihydro-4H-chromen-4-one	Cc1c(cc2c(c1O)C(=O)[C@H]([C@@H](O2)c3c cc(c(c3)O)O)O)O
4.	C ₁₆ H ₁₄ O ₇	(2R,3R)-2-(3,4-Dihydroxyphenyl)-3,5,7- trihydroxy-6-methyl-2,3-dihydro-4H- chromen-4-one	Cc1c(cc2c(c1O)C(=O)[C@@H]([C@H](O2)c3c cc(c(c3)O)O)O)O
5.	C ₁₅ H ₁₂ O ₈	(2R,3S)-3,5,7-Trihydroxy-2-(3,4,5-trihydroxyphenyl)-2,3-dihydro-4H-chromen-4-one	c1c(cc(c(c1O)O)O)[C@@H]2[C@@H](C(=O)c3 c(cc(cc3O2)O)O)O
6.	C ₁₅ H ₁₂ O ₈	(2R,3R)-3,5,7-Trihydroxy-2-(3,4,5-trihydroxyphenyl)-2,3-dihydro-4H-chromen-4-one	c1c(cc(c(c1O)O)O)[C@@H]2[C@H](C(=O)c3c(cc(cc3O2)O)O)O
7.	C ₁₅ H ₁₄ O ₆	(2R,3S)-2-(3,4-Dihydroxyphenyl)-3,5,7-chromanetriol	c1cc(ccc1[C@@H]2[C@H](Cc3c(cc(cc3O2)O) O)O)O)O
8.	C ₁₅ H ₁₄ O ₆	(2S,3R)-2-(3,4-Dihydroxyphenyl)-3,5,7-chromanetriol	c1cc(c(cc1[C@H]2[C@@H](Cc3c(cc(cc3O2)O) O)O)O)O
9.	C ₁₅ H ₁₄ O ₆	(2S,3S)-2-(3,4-Dihydroxyphenyl)-3,5,7-chromanetriol	c1cc(c(cc1[C@H]2[C@H](Cc3c(cc(cc3O2)O)O) O)O)O
10.	C ₁₅ H ₁₄ O ₆	(2R,3R)-2-(3,4-Dihydroxyphenyl)-3,5,7-chromanetriol	c1cc(c(cc1[C@@H]2[C@@H](Cc3c(cc(cc3O2) O)O)O)O)O

ADME and Toxicity Predictions

The ADME analysis revealed that astilbin's similar compounds have high gastrointestinal absorption rate and moderate solubility. Toxicity predictions indicated a low risk of hepatotoxicity, cardiotoxicity, and mutagenicity, supporting its potential as a safe therapeutic agent.

For instance amoung the 10 top structurally similar compounds only one compound with the lowest toxicity was selected for the further study and who's ADMET data is presented below

Table 2: ADME and Toxicity Profile of the least toxic molecule from top similar compounds Compound No. 2. $C_{15}H_{12}O_7$

 $(2R,\!3R)\text{-}2\text{-}(3,\!4\text{-}Dihydroxyphenyl})\text{-}3,\!5,\!7\text{-}trihydroxy\text{-}2,\!3\text{-}dihydro\text{-}4H\text{-}chromen\text{-}4\text{-}one$

PHYSICOCHEMICAL PROPERTY

Molecular Weight (MW)	304.06
Volume	285.403
Density	1.065
nHA	7.0
nHD	5.0
nRot	1.0

http://www.veterinaria.org

Article Received: Revised: Accepted:



nRing	3.0
MaxRing	10.0
nHet	7.0
fChar	0.0
nRig	18.0
Flexibility	0.056
Stereo Centers	2.0
TPSA	127.45
logS	-3.331
logP	0.932
logD7.4	1.224
pka (Acid)	7.266
pka (Base)	3.56
Melting point	248.755
Boiling point	352.509

MEDICINAL CHEMISTRY

QED	0.501
SAscore	Easy
GASA	Easy
Fsp ³	0.133
MCE-18	61.471
NPscore	2.298
Lipinski Rule	Accepted
Pfizer Rule	Accepted
GSK Rule	Accepted
GoldenTriangle	Accepted
PAINS	1
Alarm_NMR Rule	3
BMS Rule	1
Chelating Rule	1
Colloidal aggregators	0.649
FLuc inhibitors	0.535
Blue fluorescence	0.18
Green fluorescence	0.153
Reactive compounds	0.583
Promiscuous compounds	0.316

ABSPORPTION

Caco-2 Permeability	-6.151
MDCK Permeability	-4.921
PAMPA	+
Pgp inhibitor	
Pgp substrate	
HIA	

http://www.veterinaria.org

Article Received: Revised: Accepted:



F20%	++
F30%	+++
F50%	+++

DISTRIBUTION

PPB	95.1%
VDss	-0.352
BBB	
Fu	4.4%
OATP1B1 inhibitor	+++
OATP1B3 inhibitor	+++
BCRP inhibitor	++
MRP1 inhibitor	++
BSEP inhibitor	

METABOLISM

CYP1A2 inhibitor	
CYP1A2 substrate	+++
CYP2C19 inhibitor	
CYP2C19 substrate	
CYP2C9 inhibitor	
CYP2C9 substrate	
CYP2D6 inhibitor	
CYP2D6 substrate	
CYP3A4 inhibitor	+++
CYP3A4 substrate	
CYP2B6 inhibitor	
CYP2B6 substrate	
CYP2C8 inhibitor	+++
HLM Stability	

EXCRETION

CL _{plasma}	13.376
T1/2	2.089

TOXICOPHORE RULES

Acute Aquatic Toxicity	Rule	2
Genotoxic Mutagenicity Rule	Carcinogenicity	0
NonGenotoxic Carcinog	genicity Rule	0
Skin Sensitization Rule		10
Aquatic Toxicity Rule		0
NonBiodegradable		1
SureChEMBL Rule		0
FAF-Drugs4 Rule		2

TOXICITY

http://www.veterinaria.org

Article Received: Revised: Accepted:



hERG Blockers	0.061
hERG Blockers (10um)	0.538
DILI	0.472
AMES Toxicity	0.653
Rat Oral Acute Toxicity	0.3
FDAMDD	0.632
Skin Sensitization	0.986
Carcinogenicity	0.241
Eye Corrosion	0.017
Eye Irritation	0.984
Respiratory	0.671
Human Hepatotoxicity	0.499
D 1 N 1	
Drug-induced Nephrotoxicity	0.126
	0.126 0.037
Drug-induced Neurotoxicity	0.037
Drug-induced Neurotoxicity Ototoxicity	0.037 0.675
Drug-induced Neurotoxicity Ototoxicity Hematotoxicity	0.037 0.675 0.078
Drug-induced Neurotoxicity Ototoxicity Hematotoxicity Genotoxicity	0.037 0.675 0.078 0.992
Drug-induced Neurotoxicity Ototoxicity Hematotoxicity Genotoxicity RPMI-8226 Immunitoxicity	0.037 0.675 0.078 0.992 0.05
Drug-induced Neurotoxicity Ototoxicity Hematotoxicity Genotoxicity RPMI-8226 Immunitoxicity A549 Cytotoxicity	0.037 0.675 0.078 0.992 0.05 0.769
Drug-induced Neurotoxicity Ototoxicity Hematotoxicity Genotoxicity RPMI-8226 Immunitoxicity A549 Cytotoxicity Hek293 Cytotoxicity	0.037 0.675 0.078 0.992 0.05 0.769 0.582
Drug-induced Neurotoxicity Ototoxicity Hematotoxicity Genotoxicity RPMI-8226 Immunitoxicity A549 Cytotoxicity Hek293 Cytotoxicity BCF	0.037 0.675 0.078 0.992 0.05 0.769 0.582 0.998

TOX21 PATHWAY

NR-AhR	++
NR-AR	
NR-AR-LBD	
NR-Aromatase	
NR-ER	
NR-ER-LBD	+
NR-PPAR-gamma	
SR-ARE	+
SR-ARE SR-ATAD5	+
	+
SR-ATAD5	

Cytoscape Network Analysis

The Cytoscape network analysis identified key proteins and pathways involved in oxidative stress and antioxidant defense mechanisms. Significant interactions were observed with enzymes such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx).

http://www.veterinaria.org

Article Received: Revised: Accepted:



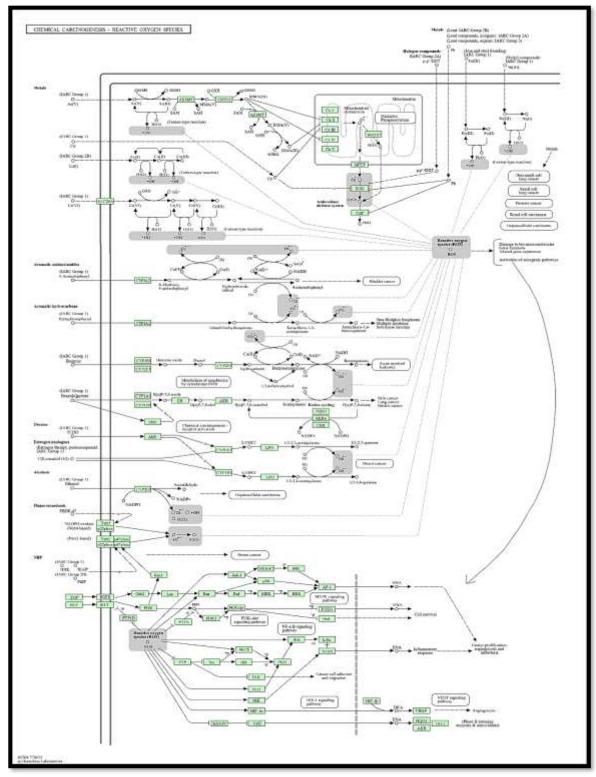


Figure 1: Cytoscape Network Analysis of Protein-Protein Interactions for antioxidant activity

Molecular Docking Studies

Molecular docking studies demonstrated that the compound $C_{15}H_{12}O_7$ has strong binding affinities to key antioxidant enzymes. For Human Erythrocyte Catalase (PDB ID: 1DGF), the compound $C_{15}H_{12}O_7$ exhibited a docking score of -28.20624, while ascorbic acid had a docking score of -22.32259. For Human Peroxiredoxin 3 (PDB ID: 5UCX), $C_{15}H_{12}O_7$ had a docking score of 21.29366 compared to -12.70578 for ascorbic acid.

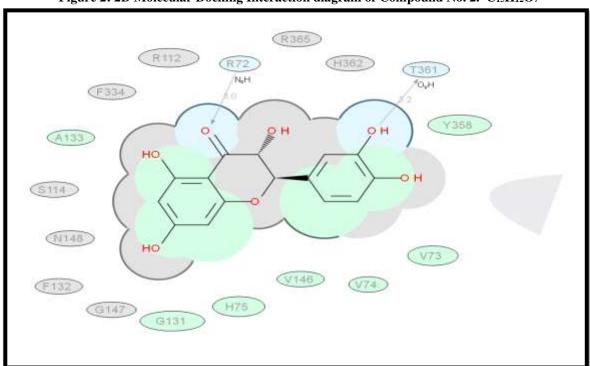
Article Received: Revised: Accepted:



Table 3: Docking Scores of Compound No. 2 C₁₅H₁₂O₇ and Ascorbic Acid.

Enzyme	Compound	Docking Score
Human Erythrocyte CAT	$C_{15}H_{12}O_7$	-28.20624
Human Erythrocyte CAT	Ascorbic Acid	-22.32259
Human Peroxiredoxin 3	$C_{15}H_{12}O_7$	-21.29366
Human Peroxiredoxin 3	Ascorbic Acid	-12.70578

Figure 2: 2D Molecular Docking Interaction diagram of Compound No. 2. C₁₅H₁₂O₇



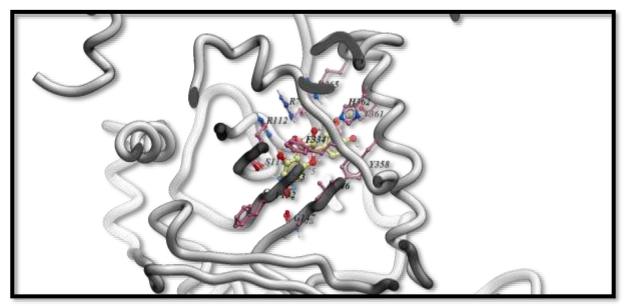


Figure 3: 2D Molecular Docking Interaction diagram of Compound No. 2. C₁₅H₁₂O₇ with Human Erythrocyte protein.

http://www.veterinaria.org

Article Received: Revised: Accepted:



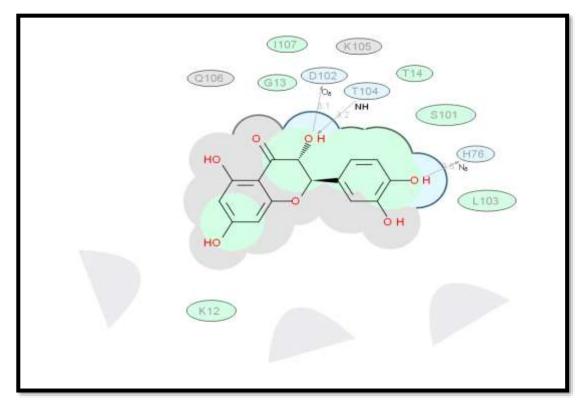


Figure 4: 2D Molecular Docking Interaction diagram of Compound No. 2. C₁₅H₁₂O₇ with Human Peroxidin 3 protein.

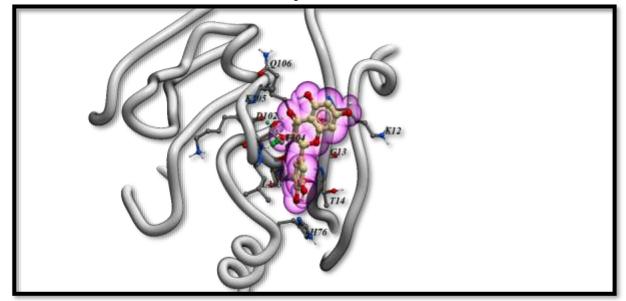


Figure 5: 3D Molecular Docking Interaction diagram of Compound No. 2. C₁₅H₁₂O₇ with Human Peroxidin 3 protein.

Discussion

The comprehensive in-silico analysis of astilbin related similar compounds underscores its potential as a potent antioxidant agent. The Swiss Similarity studies provided a robust framework for identifying structurally similar compounds, while ADME and toxicity predictions confirmed its favourable pharmacokinetic and safety profiles. Cytoscape network analysis elucidated the complex interactions within antioxidant pathways, and molecular docking studies validated astilbin's strong binding affinities to critical enzymes involved in oxidative stress defense. Comparative analysis with ascorbic acid further underscores astilbin's therapeutic potential.

Conclusion

REDVET - Revista electrónica de Veterinaria - ISSN 1695-7504

Vol 24, No. 4 (2023)

http://www.veterinaria.org

Article Received: Revised: Accepted:



Astilbin, derived from the rhizomes of Smilax glabra, exhibits promising antioxidant properties validated through comprehensive in-silico studies. The favourable ADME profile, low toxicity, and strong binding affinities to key antioxidant enzymes position astilbin as a viable candidate for further experimental validation and potential therapeutic application in antioxidant therapy.

References

- 1. Swiss Institute of Bioinformatics. Swiss Similarity: Web tool for rapid structural comparison of bioactive molecules [Internet]. Available from: https://www.swisssimilarity.ch/
- 2. Gfeller D, Grosdidier A, Wirth M, Daina A, Michielin O, Zoete V. SwissTargetPrediction: A web server for target prediction of bioactive small molecules. Nucleic Acids Res. 2014 Jul;42(Web Server issue)
- 3. Daina A, Michielin O, Zoete V. SwissADME: A free web tool to evaluate pharmacokinetics, drug-likeness, and medicinal chemistry friendliness of small molecules. Sci Rep. 2017 Mar 3;7:42717.
- 4. Mendez D, Gaulton A, Bento AP, Chambers J, De Veij M, Félix E, et al. ChEMBL: Towards direct deposition of bioassay data. Nucleic Acids Res. 2019 Jan 8;47(D1)
- 5. Irwin JJ, Sterling T, Mysinger MM, Bolstad ES, Coleman RG. ZINC: A free tool to discover chemistry for biology. J Chem Inf Model. 2012 Jul 23;52(7):1757-68.
- 6. Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, et al. DrugBank 5.0: A major update to the DrugBank database for 2018. Nucleic Acids Res. 2018 Jan 4;46(D1)
- 7. BindingDB. BindingDB: The Binding Database. Available from: https://www.bindingdb.org/. Accessed 17/05/2024.
- 8. ADMET LAB 3.0. ADMET LAB 3.0: Predictive modeling for drug absorption, distribution, metabolism, excretion, and toxicity. Available from: https://admetlab3.scbdd.com/. Accessed 21/05/2024.
- 9. STRING: STRING: Functional protein association networks [Internet]. Available from: https://string-db.org/. Accessed 25/05/2024.
- 10. Cytoscape. Cytoscape: An Open-Source Platform for Complex Network Analysis and Visualization [Internet]. Available from: https://cytoscape.org/. Accessed 26/05/2024.
- 11. KEGG. Kyoto Encyclopedia of Genes and Genomes (KEGG) [Internet]. Available from: https://www.genome.jp/kegg/. Accessed 27/05/2024.
- 12. Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, et al. STRING v11: protein–protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 2019;47(D1) Available from: https://academic.oup.com/nar/article/47/D1/D607/5198476. Accessed 28/05/2024.
- 13. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. GenomeRes. 2003;13(11):2498-504. Available from: https://genome.cshlp.org/content/13/11/2498.short. Accessed 29/05/2024.
- 14. Kanehisa M, Goto S, Sato Y, Furumichi M, Tanabe M. KEGG for integration and interpretation of large-scale molecular data sets. Nucleic Acids Res. 2012;40(D1) Available from: https://academic.oup.com/nar/article/40/D1/D109/2904084. Accessed 30/05/2024.
- 15. Bader GD, Cary MP, Sander C. Pathguide: a pathway resource list. Nucleic Acids Res. 2006;34(suppl_1)Availablefrom: https://academic.oup.com/nar/article/34/suppl_1/D504/1130207.Accessed 1/05/2024.
- 16. Cline MS, Smoot M, Cerami E, Kuchinsky A, Landys N, Workman C, et al. Integration of biological networks and gene expression data using Cytoscape. Nat Protoc. 2007;2(10):2366-82. Available from: https://www.nature.com/articles/nprot.2007.324. Accessed 28/05/2024.
- 17. MolSoft L.L.C. ICM-Pro User Manual. Available from: https://www.molsoft.com/manual/index.html
- 18. Morris GM, Lim-Wilby M. Molecular docking. Methods Mol Biol. 2008;443:365-82.
- 19. Kirkpatrick S, Gelatt CD, Vecchi MP. Optimization by simulated annealing. Science. 1983;220(4598):671-80.
- 20. Halperin I, Ma B, Wolfson H, Nussinov R. Principles of docking: An overview of search algorithms and a guide to scoring functions. Proteins. 2002;47(4):409-43.
- 21. Kitchen DB, Decornez H, Furr JR, Bajorath J. Docking and scoring in virtual screening for drug discovery: methods and applications. Nat Rev Drug Discov. 2004;3(11):935-49.